

NASTASSIA VIRGINIA PATIN
www.nastassiavpatin.com

Cooperative Institute for Marine and Atmospheric Studies
Rosenstiel School of Marine and Atmospheric Sciences
University of Miami, Miami, FL

650 521 3888 (mobile)
nastassia.patin@noaa.gov

Ocean Chemistry & Ecosystems Division
Atlantic Oceanographic and Meteorological Laboratory
Oceanic and Atmospheric Research
National Oceanic and Atmospheric Administration
Stationed at Southwest Fisheries Science Center, La Jolla, CA

RESEARCH INTERESTS

Marine microbial ecology, genomics and metagenomics, bioinformatics, marine food webs

EDUCATION AND PROFESSIONAL EXPERIENCE

National Oceanographic and Atmospheric Administration
Postdoctoral Researcher at the University of Miami/AOML
Advisor: Kelly Goodwin
Aug. 2020 - present

Georgia Institute of Technology
Postdoctoral researcher
Advisor: Frank Stewart
Co-advisor: Kostas Konstantinidis (as of February 2019)
2016 – Aug. 2020

Scripps Institution of Oceanography, University of California, San Diego
Ph.D. in Marine Biology
Advisor: Paul Jensen
2011 – 2016

San Francisco State University
M.S. in Marine Biology
Advisor: Frances Wilkerson
Co-advisor: Matthew Ashby
2009 – 2011

Stanford University
B.S. in Biological Sciences, minor in French.
2004 –2008

PUBLICATIONS

Patin NV, Goodwin KD. Capturing Marine Microbiomes: A field sampling guide. *In preparation for Frontiers in Marine Microbiology.*

Patin NV, Jesser KJ, Peña-Gonzalez A, Hatt JK, Trueba G, Levy K, Konstantinidis KT. Comparison of metagenomic and traditional methods for diagnosis of *E. coli* enteric infections. *Under review at mBio.*

Truelove N, **Patin NV**, Min M, Pitz K, Chavez F, Goodwin K. Expanding the scale of environmental DNA research with autonomous sampling. *Under revisions for eDNA.*

- Thompson LR, Anderson SR, Den Uyl PA, **Patin NV**, Sanderson G, Goodwin KD. Tourmaline: a workflow for rapid and reproducible amplicon sequence analysis using QIIME 2 and Snakemake. *Under revisions for GigaScience*.
- Demko A, **Patin NV**, Jensen PR. 2021. Culture-dependent and culture-independent microbial diversity in tropical marine sediments. *Environmental Microbiology*. 23(11), 6859–6875. Doi: 10.1111/1462-2920.15798
- Caughman A, Pratte ZA, **Patin NV**, Stewart PJ. 2021. Coral microbiome changes over the day-night cycle. *Coral Reefs*. <https://doi.org/10.1007/s00338-021-02097-8>
- Patin NV**, Dietrich ZA, Stancil A, Quinan M, Beckler JS, Hall ER, Culter J, Smith CG, Taillefert M, Stewart FJ. 2021. Gulf of Mexico blue hole harbors high levels of novel microbial lineages. *The ISME Journal*. doi: s41396-021-00917-x
- Abdelrahman SM, **Patin NV**, Hanora AM, Aboseidah AA, Desoky SM, Desoky FG, Stewart FJ, Lopanik NB. 2021. The natural product biosynthetic potential of Red Sea nudibranch microbiomes. *PeerJ* 9:e10525. doi: 10.7717/peerj.10525
- Patin NV**, Peña-Gonzalez A, Hatt J, Moe C, Kirby A, Konstantinidis K. 2020. The role of the gut microbiome in resisting norovirus infection as revealed by a human challenge study. *mBio*. 11 (6) e02634-20. doi: 10.1128/mBio.02634-20.
- Patin NV**, Brown E, Garfield C, Chebli G, Kubanek J, Stewart FJ. 2020. Microbial and chemical dynamics of a toxic dinoflagellate bloom. *PeerJ*. 8:e9493. doi: 10.7717/peerj.9493
- Thamdrup B, Steinsdottir HGR, Bertagnolli AD, Padilla CC, **Patin NV**, Garcia-Robledo E, Bristow L, Stewart FJ. 2018. Anaerobic methane oxidation is an important sink for methane in the ocean's largest oxygen minimum zone. *Limnology and Oceanography* 9999: 1-17. doi: 10.1002/lno.11235
- Patin NV**, Locklear S, Stewart FJ, Lopanik NB. 2018. Symbiont frequency predicts microbiome composition in a model bacterial-bryozoan symbiosis. *Aquatic Microbial Ecology* 83: 1-13. doi:10.3354/ame01901
- Tuttle RN, Demko A, **Patin NV**, Kapon C, Dorrestein PC, Jensen PR. 2018. The detection of specialized metabolites and their producers in ocean sediments. *Applied and Environmental Microbiology*. doi:10.1128/AEM.02830-18
- Pratte ZA, **Patin NV**, McWhirt M, Caughman A, Stewart FJ. 2018. Association with a sea anemone alters the skin microbiome of clownfish. *Coral Reefs*. doi:10.1007/s00338-018-01750-z
- Patin NV**, Floros D, Dorrestein PC, Hughes C, Jensen PR. 2018. The role of inter-species interactions in *Salinispora* specialized metabolite production. *Microbiology*. 164(7): 946-955.
- Patin NV**, Pratte ZA, Regensburger M, Gilde K, Hall E, Dove ADM, Stewart FJ. 2018. Microbiome dynamics in a large artificial seawater aquarium. *Applied and Environmental Microbiology*. 84(10): e00179-18.
- Patin NV**, Schorn MA, Aguinaldo K, Lincecum T, Moore BS, Jensen PR. 2016. Effects of actinomycete secondary metabolites on sediment microbial communities. *Applied and Environmental Microbiology*. 83(4): e02676-16.

Schorn MA, Alanjary MM, Aguinaldo K, Korobeynikov A, Podell S, **Patin NV**, Lincecum T, Jensen PR, Ziemert N, Moore BS. 2016. Sequencing rare marine actinomycete genomes reveals high density of unique natural product biosynthetic gene clusters. *Microbiology*. 162(12): 2075-2086.

Patin NV, Duncan K, Dorrestein PC, Jensen PR. 2016. Competitive strategies differentiate closely related species of marine actinobacteria. *The ISME Journal*. 10: 478-490.

Wietz M, Duncan K, **Patin NV**, Jensen PR. 2013. Antagonistic interactions mediated by marine bacteria: The role of small molecules. *Journal of Chemical Ecology*. 9: 879-891.

Patin NV, Kunin V, Lidström U, Ashby M. 2012. Effects of OTU clustering and PCR artifacts on microbial diversity estimates. *Microbial Ecology*. 65(3): 709-19.

Bagulayan A, Bartlett-Roa JN, Carter AL, Inman BG, Keen EM, Orenstein EC, **Patin NV**, Sato KNS, Sibert EC, Simonis AE, Van Cise AM, Franks PJS. 2012. Journey to the center of the gyre: The fate of the Tohoku Tsunami debris field. *Oceanography* 25(2): 200–207.

CURRENT RESEARCH PROJECTS

Comparing marine microbial communities in the California Current sampled by an autonomous environmental sampler compared to manual bottle sampling using shotgun metagenomics.

Characterizing California Current microbiomes over temporal, spatial, and oceanographic scales using shotgun metagenomics.

Linking microbes to higher organisms and oceanographic conditions using eDNA and 'Omics.

AWARDS, HONORS, AND RESEARCH SUPPORT

Peggy Cotter Travel Award, American Society for Microbiology	2019
Best Talk, Georgia Tech Postdoctoral Symposium	2017
Edward A. Frieman Director's Prize for Excellence in Graduate Student Research <i>Awarded for the 2016 ISMEJ publication listed above.</i>	2016
International Society for Microbial Ecology Student Travel Grant	2014, 2016
SIO Department Graduate Student Excellence Travel Award	2014, 2016
San Francisco Bay Scholarship	2009
DAAD Research Internship in Science and Engineering Fellowship	2007
Stanford Undergraduate Research Grant	2006

PRESENTATIONS AND INVITED TALKS

Patin NV , Pitz K, Truelove N, Chavez F, Goodwin K. Shotgun metagenomes combined with metabarcoding link microbial function to higher trophic levels in the California Current. Oral	2021
---	------

presentation at the 5th annual eDNA Technical Exchange Workshop.

Patin NV, Pitz K, Truelove N, Chavez F, Goodwin K. Comparing microbial eDNA captured by automated and manual sampling methods. Oral presentation at NCCOS-AOML Symposium. 2021

Patin NV, Dietrich Z, Beckler J, Hall E, Stewart FJ. Metagenome-assembled genomes of the rare and enigmatic Woese archaeota inform their role in blue hole biogeochemistry. Oral presentation at Ocean Sciences Meeting 2020. 2020

Patin NV, Brown E, Chebli G, Garfield C, Kubanek J, Stewart FJ. Microbial and chemical dynamics of a toxic dinoflagellate bloom. Oral presentation at 10th US Symposium on Harmful Algae. 2019

Patin NV, Pratte ZA, Regensburger M, Gilde K, Hall E, Dove ADM, Stewart FJ. The *Ocean Voyager* Microbiome. Poster. American Society for Microbiology Microbe 2019 Conference. 2019

Patin NV. Chemical Ecology in the Age of Microbiome Science and Big Data. **Invited Talk**. 35th annual meeting of the International Society of Chemical Ecology. Atlanta, GA. 2019

Patin NV, Pratte ZA, Regensburger M, Gilde K, Hall E, Dove ADM, Stewart FJ. The *Ocean Voyager* Microbiome. **Invited Talk**. Kennesaw State University. 2018

Patin NV, Pratte ZA, Regensburger M, Gilde K, Hall E, Dove ADM, Stewart FJ. The *Ocean Voyager* Microbiome. **Invited Talk**. Astrobiology Graduate Conference, Georgia Institute of Technology. 2018

Patin NV, Locklear S, Stewart FJ, Lopanik NB. The microbiome of the bryozoan *Bugula neritina* is shaped by a cytotoxin-producing symbiont. Poster. 7th Conference on Beneficial Microbes. 2018

Patin NV, Pratte ZA, Regensburger M, Gilde K, Hall E, Dove ADM, Stewart FJ. The *Ocean Voyager* Microbiome. Poster. 11th Georgia Tech Bioinformatic Conference. 2017

Patin NV, Pratte ZA, Regensburger M, Gilde K, Hall E, Dove ADM, Stewart FJ. The *Ocean Voyager* Microbiome. Talk. Georgia Tech Postdoctoral Symposium. 2017

Patin NV, Schorn M, Aguinaldo K, Lincecum T, Moore BS, Jensen PR. Effects of actinomycete secondary metabolites on sediment microbial communities. Poster. International Society for Microbial Ecology-16 Conference. 2016

Patin NV, Duncan K, Dorrestein PC, Jensen PR. Competitive strategies differentiate closely related species of marine actinobacteria. Poster. International Society for Microbial Ecology-15 Conference. 2014

Patin NV, Kunin V, Lidström U, Ashby M. Effects of OTU clustering and PCR artifacts on microbial diversity estimates. Poster. International Society for Microbial Ecology-13 Conference. 2010

SERVICE, OUTREACH, AND MEDIA

Invited Member, NOAA 'Omics Working Group Subcommittee on bioinformatics and data management	07/2021
Author, <i>It's the little things: Studying marine microbes.</i> Blog post: Cooperative Institute for Marine and Atmospheric Sciences, Rosenstiel School For Marine and Atmospheric Sciences. https://cimas.rsmas.miami.edu/news-events/cimas-blog/little-things/index.html	02/2021
Author, <i>Diving for microbes: journey into a blue hole.</i> Nature Microbiology: Behind the Paper. https://naturemicrobiologycommunity.nature.com/posts/diving-for-microbes-journey-into-a-blue-hole	02/2021
Founder and Organizer, Georgia Tech Bioinformatics User Group (GT-BUG) Established a regular meeting of students, postdoc, and faculty interested in bioinformatics for discussions, tutorials, and invited speakers in order to address community needs in sharing knowledge and experience among scientists from Georgia Tech, Emory University, the Centers for Disease Control and Prevention, and Georgia State University.	2017-present
Co-organizer, Summer Workshop in Marine Science (SWiMS) Helped developed tutorials on incorporating marine science into high school lesson plans and delivered tutorials to workshop attendees (high school teachers).	2017, 2018
Visiting speaker, Gwinnett High School. Spent a day giving short talks and interactive lessons on ocean pollution to 9 th graders.	2018
Judge, Morningside Elementary School Science Fair Assessed and ranked the quality of 3 rd -5 th grade science fair projects.	2017, 2018
Visiting speaker, Northwestern Middle School Presented short talks and interactive lessons on microbiology to 6 th and 7 th graders.	March 2017
Co-coordinator, Scripps Community Outreach Program for Education Facilitated educational outreach opportunities at SIO and volunteered for these opportunities.	2013 – 2016
Student Representative, SIO Heritage Committee Generated recommendations to the UC San Diego administration for the upkeep and preservation of historical buildings, facilities, and other resources.	2015 - 2016
Organizer, Microbial Oceanography Journal Club Organized monthly paper discussions for graduate students and postdocs.	2012 –2014
Member, Marine Biology Curricular Group Student Committee Wrote evaluations for SIO faculty members based on course evaluations, instructor evaluations, and student feedback.	2012, 2013
Volunteer, Sea Lion Bowl, National Ocean Sciences Bowl Assisted with preparation and logistics of two annual high school marine science quiz bowls.	2009, 2010

FIELD EXPERIENCE

Mote Marine Lab, Sarasota, Florida Led the microbial sampling of blue hole formations in the Gulf of Mexico over three field expeditions. Collected water and sediment samples via SCUBA diving and robotic lander deployment, in collaboration with Mote Marine Laboratory and Florida Atlantic University. Two expeditions sampling included mentorship of students who gained valuable field experience and one of whom is a co-author the resulting publication.	2019, 2020
Mote Marine Lab, Sarasota, Florida Organized and led a field collection experiment to the southwest coast of Florida with two undergraduate students. Collected water samples from twelve sites and size-fractionated for particle-associated and free-living microbial community analyses. Taught all methods to the accompanying students and included them as co-authors on a pending publication.	2018
Gump Research Station, Mo'orea, French Polynesia Collected crabs for gut microbiome analyses. Conducted feeding experiments on crabs for gut transcriptome analyses. Collected seaweeds for study on nutrient loads and algal microbiomes. Sampled coral over a 48-hour period for diel microbiome study.	2017
Institute of Marine Sciences, Morehead City, North Carolina Collected colonies of the bryozoan <i>Bugula neritina</i> for a study on microbiome structure, function, and biogeography. Assisted with ecological experiments assessing predation levels in different <i>B. neritina</i> habitats.	2017
Carrie Bow Cay Field Station, Belize Conducted <i>in situ</i> experiments to detect microbial bioactive compounds in marine sediments and to assess effects of actinomycete compounds on natural communities.	2014, 2015
Viti Levu, Fiji Conducted <i>in situ</i> experiments on microbial chemical competition. Collected sediment samples for cultivation of actinomycete bacteria.	2014
UNOLS cruise, Bahamas Collected sediment samples for cultivation of actinomycete bacteria and culture-independent sediment community analyses.	2013
UNOLS cruise, Yucatán Peninsula, Mexico Collected sediment, algal, and invertebrate samples for cultivation of actinomycete bacteria.	2012

TEACHING AND MENTORING EXPERIENCE

Mentor , Zoë Dietrich (summer REU student, fall field assistant) Guided student through sample processing and analysis of marine water column and sediment samples obtained from a blue hole field expedition. By the end of the summer the student had produced high-quality amplicon sequence data and interpreted the results in oral presentation, poster, and written report formats. The student participated in the second sampling trip and contributed substantially	summer 2019
---	-------------

to sample collection and processing. She is anticipated to be a co-author on future publications describing our results.

Mentor, Chloe Pryor (undergraduate) fall 2018-present
Taught molecular biology laboratory skills to a first-year undergraduate, leading to the generation of high-throughput sequencing data set. Currently teaching basic bioinformatic data analysis methods (quality control, taxonomic assignment, and ecological statistics).

Mentor, Claire Garfield and Gabriella Chebli (summer REU students) summer 2018
Developed a summer research project incorporating field collections, molecular biology, and metabolomic data generation and analyses for two undergraduate students. Guided students through all components of the project including a 4-day field trip to Florida as the single head scientist in charge of all aspects of the trip (scientific, educational, and logistical). By the end of the summer students had generated two major preliminary data sets that were used in an NSF grant proposal. They also presented their results as a poster and a paper at the Georgia Tech REU Research Symposium.

Organizer/lecturer, Georgia Tech Bioinformatics User Group 2017 - present
Conducting tutorials on basic bioinformatic programs and skills on topics including amplicon and metagenome sequencing data analysis, phylogenetics, and ecological statistics.

Mentor, Doug Sweeney (undergraduate) 2015 - 2016
Taught microbiology and molecular biology techniques to an undergraduate student, leading to an independent research project and his ultimately joining the lab as a PhD student.

Teaching Assistant, SIO282: Microbial Life in Extreme Environments, UCSD 2016
Prof. Doug Bartlett
Graded exams and conducted exam review sessions. Designed and presented a lecture on microbial symbiosis.

Teaching Assistant, GES56: Changes in the Coastal Ocean, Stanford, CA 2008
Prof. Rob Dunbar
Provided teaching support for class on the science and policy of the California coast. Created and executed lesson plan about the Channel Islands marine reserve.

PROFESSIONAL ASSOCIATIONS

American Association for the Advancement of Science
The American Society of Microbiology

OTHER

Citizenship: United States and Switzerland
Languages: Proficient in French and German, some knowledge of Spanish
Society Memberships: Union of Concerned Scientists, 500 Women Scientists, Sierra Club, Surfrider Foundation
Other: AAUS Scientific Diver, PADI Divemaster